

StabilCoat®

Immunoassay Stabilizer

Procedures

Store at 4 °C. Use straight or dilute 1:1 in deionized water or PBS. No preservatives added. If you do not plan to use within 7 days after opening, use aseptic technique when removing contents, or add a preservative compatible with your assay.

To Stabilize Adsorbed or Immobilized Proteins on Microwell Plates/Strips

1. Immobilize or adsorb the primary protein (antibody or antigen) according to the procedure optimized in your laboratory. Wash adequately to remove excess or weakly bound protein.
2. Immediately after washing, add StabilCoat solution to allow interaction with the entire protein-coated surface. For example, if you added 100 µl/well of the primary protein solution in step one, then add 100 µl/well StabilCoat solution. Do not let coated components dry before adding StabilCoat solution since drying contributes to the loss of protein activity.
3. Incubate for 15 to 60 minutes at room temperature. For most assays, StabilCoat can replace the blocking solution. However, if your assay demands more blocking, mix StabilCoat 1:1 with your current blocking solution for added blocking capability.
4. Remove or aspirate the StabilCoat solution, but do not wash.
5. Dry components for long-term storage. Products coated with StabilCoat may require longer drying times than those without StabilCoat. Recommended methods are:
 - place plates in a humidity controlled chamber (less than 15% humidity) until dry (4 to 24 hours).
 - or dry plates at 30-40°C in a vacuum oven for 4 hours.

Drying times should be optimized for each application.

6. Package the final, stabilized product in an airtight container with a desiccant. This is especially important when the final product is stored in a humid environment or refrigerated (where condensation is likely to occur).

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StabilCoat®

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To Stabilize Adsorbed or Immobilized Proteins on Membranes

Following are general guidelines only. Additional optimization of concentrations and application method may be required to meet the shelf life and flow requirements for each assay.

1. Immobilize or adsorb the primary protein (antibody or antigen) according to the procedure optimized in your laboratory.
2. Dilute 1 part StabilCoat in 3 parts compatible buffer or deionized water.
3. Add 0.01% surfactant.
4. Coat membrane by incubating or spraying with the StabilCoat mixture.
5. Dry thoroughly. Faster drying results in better flow properties.
6. Package the final, stabilized product in an airtight container with a desiccant. This is especially important when the final product is stored in a humid environment or refrigerated (where condensation is likely to occur).

To Stabilize Conjugates in Dry Form

Following are recommended guidelines for stabilizing conjugates in a dried form:

1. If you dilute your conjugate before drying/lyophilizing, use StabilCoat as the diluent. Otherwise, add between 5 and 10 parts StabilCoat to 1 part conjugate. (Optimum ratio should be empirically determined.) Mix gently.
2. If you are lyophilizing in vials with rubber stoppers, stability can sometimes be improved by placing the rubber stoppers in a 100°C vacuum oven for one hour before use. This dries them and drives off any volatiles present in the stopper.
3. Freeze the conjugate/stabilizer mixture, then lyophilize as normal. Lyophilization may require extra time.
4. For evaporation drying, place the conjugate/stabilizer mixture in a 37-40°C oven for 4 hours or until completely dry. The volume per container should be low enough to allow maximum surface area to be exposed to air during drying. Store the final product in an airtight container.

For technical assistance call (760) 806-8922

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